

A NEW METHOD OF CHALKBROOD CONTROL USING CHITOSAL-APIS[®] LIQUID

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S u m m a r y

Most known antifungals act on enzymes involved in the biosynthesis of ergosterol in fungi. Chitin-derivatives, potent antifungal preparations most probably act on the permeability of membranes of fungal hypha. Chitosal-Apis[®] Liquid which is a mixture of 2-acetamido-2-deoxy- β -D-glucose and 2-amino-deoxy- β -glucopyranose in 1% salicylic acid not only inhibits but also kills *Ascosphaera apis* – a causative agent of chalkbrood of the honey bee. It is virtually non toxic for larval brood and worker bees after oral administration. Both spring and autumn treatments with Chitosal-Apis[®] Liquid were very effective. No side effects were observed during and after the therapy. Chitosal-Apis[®] Liquid is a natural product and therefore not dangerous either to bees or to consumers of the honey bee products.

Keywords: Chitosal Apis[®] Liquid, chalkbrood, honey bee.

INTRODUCTION

Among infectious diseases of the honey bee reported in 1999 fungal infections appear to be prevalent in many countries (Shimanuki and Knox 1999) (Tab.1). Fungi are not only common saprophytes of bees and combs. Most of the fungi collected by bees are pathogenic (Gliński et al. 1988) (Tab.2). In the honey bee fungi initiate infection by spores germination. The developing invasive hypha penetrates the cuticle mechanically and enzymatically enters the bee's body cavity where it rapidly develops and overwhelms the internal organs. The invasion may sometimes start from ingested fungal spores that germinate in the intestines. Physical, chemical and biological stress factors, mainly extremal temperatures and high humidity, environmental pollution, pesticide poisonings, parasite invasions and attacks of predators are the factors which lead to the development of fungal infection in bees. Hence, stress conditions are the risk factors because they reduce the resistance pattern of the insect organism by compromising the immune system and by impairing the protective barriers of the body coverings, alimentary tract and tracheae. Immunologically compromised brood of the honey bee is very sensitive to *A. apis* infection. (Stark and Gliński 1996). In the chalkbrood, infected larvae *Ascosphaera apis* both mechanically and

enzymatically damage tissues and disturb blood circulation and growing mycelium in the midgut is responsible for competition for food and place. The result of the infections is death and the mummification of the bee larvae. The fungus overgrowing the mummified bee larvae produces an enormous number of infective spores, resistant to common antifungal drugs and disinfectants. (Gliński and Chmielewski 1983)

Tab. 1

World bee health report 1999
Najczęściej występujące choroby pszczoł w wybranych krajach - 1999

Country	Chalk brood	AFB	EFB	SB	Nosema	Amoeba	Acarapis	Varroa	Tropilaelaps	Braula
Poland	+	+	+	+	+	+	+	+		+
Germany	+	+	+		+	+	+	+		+
USSR-Former	+	+	+	+	+	+	+	+	+	+
Sweden	+	+	+		+			+		+
Finland	+	+	+	+	+		?	+		+
France	+	+	+	+	+	+	+	+		+
Australia	+	+	+	+	+	+				+

Tab. 2

Mycoses of the honey bee - Grzybice pszczoły miodnej

Disease	Aetiology	Pathogenicity		Occurrence
		Brood	Imago	
Chalkbrood	<i>Ascospaera apis</i>	+		worldwide
Stonebrood	<i>Aspergillus flavus</i> <i>A. niger</i> , <i>A. fumigatus</i>	+	+	sporadic
Melanosis	<i>Areobasidium pullulans</i>		+	sporadic
Molds	<i>Trichoderma lignorum</i> , <i>T. koenigi</i> , <i>Mucor hiemalis</i>	+	+	sporadic sporadic
Yeasts	<i>Saccharomyces cerevisiae</i> , <i>S. elipsoideus</i> , <i>Kloeckera apiculata</i> , <i>Torulopsis</i> , <i>Candida</i>		+	sporadic

Only a few antifungal agents can be used to cause the selective elimination of *Ascospaera apis* from the honey bee brood with the minimal

toxicity to the brood, the bees and the queen. Drugs by contaminating honey and pollen loads are a potential danger to the consumers of these bees products. (Gliński 2000) Up to now the following have been commonly used to control chalkbrood: polyenes, (Amphotericin B, Nystatin) (Gliński and Osipowski 1984), Gryseofulvin, sorbic acid, Thiabendazol and sodium propionate. (Gliński et al. 1988). Trials have been done with Clotrimazole (Gliński et al. 1988), furanocoumarins of plant origin and scent oils (citral and geraniol). (Gliński and Chmielewski 1992, Gliński et al. 1988, Wolski et al. 1997). Very interesting results have been noted with chitosan, a mixture of derivatives obtained from crustacean chitin. Chitosan shows antifungal, bacteriostatic and antitumor properties (Wolski et al. 1992). It activates the immune system and cell regeneration in vertebrates. Recently, it was found that N-acetylated derivatives of chitin induce haemocytic and cell-free immune responses in the honey bee. (Gliński et al. 2000)

MATERIAL AND METHODS

Chitosal-Apis® Liquid is a solution of chitosan in 1% salicylic acid prepared by „Vet-Agro” Lublin. For comparison the following antimycotics were included; extracts of *Archangelica officinalis*, *Pastinaca sativa* and *Libanotis intermedia* (active substance - furanocoumarins), Nystatin and choline salt of N-glucosyl-polyfungine.

The value of minimal inhibitory concentration (MIC) and minimal fungicide concentration (MFC) were calculated by the method used by Ericsson and Sherris on Sabouraud's medium with yeast extract at 25°C; 10 isolates of *Ascosphaera apis* from Poland and three isolates from Finland (Fin.1, Fin.2, Fin.14) were tested. The following reference strains of *A. apis* were included: USDA 1, USDA 2, Uppsala 1002 and 1005.

Toxicity for brood. On brood frames of an uncaged honey bee colony, free of diseases, a test area with 50 larvae 72-80 hrs old was plotted. Using a Hamilton's microsyringe Chitosal-Apis® Liquid in 50% royal jelly (royal jelly diluted with an aqueous glucose-fructose solution (Rembold H. et al. 1974); was applied in a dose of 1.0 µ directly 1 to the larvae in the brood cells. Concentration of the dissolved test substance ranged from 10⁰ to 10² MIC for *A. apis*. The brood loss rate was also determined in an untreated control area of brood. During the test the temperature was kept at 35°C and relative humidity at 60-70%. The time limit chosen for mortality counts was 24 hours.

Toxicity for bees was determined on workers at the age of 2-3 weeks in a feeding test. The bees - in small cages in groups of 25 bees each - were allowed to feed on 25% sugar solution which contained a known amount of the test material (10⁰-10² MIC of Chitosal-Apis® Liquid for *A. Apis*) for 24 hrs. The test was performed at 30°C and 45% RH (12). The time limit chosen

for mortality counts was 24 hrs. The bees fed on the sugar solution without Chitosal® served as a control. Twenty replications were run of each test. The value of LD₅₀ was calculated according to Litchfield and Wilcoxon (Litchfield and Wilcoxon 1949) on the basis of the number of dead or viable brood or the number of bees.

Fields examinations. The therapeutic effects of Chitosal-Apis® Liquid were checked in 8 randomly chosen apiaries on 200 honey bee colonies in which chalkbrood was clinically diagnosed and then confirmed by the isolation of *Ascosphaera apis*. The level of infection varied from 25 to 50 mummies per comb. In the spring treatment Chitosal-Apis® Liquid was sprayed over brood and bees and the inside of the hive at a dose of 20 ml per colony, twice at 5 day intervals in 100 colonies. The autumn treatment was done in 100 colonies after removing the honey from the hive. The preparation at a strength of 20-25 ml per colony was sprayed over the brood, the bees and the inside of the hive twice at 5 day intervals. Fifty untreated, chalkbrood - sick honey - bee colonies served as a control.

RESULTS

Over the past few years, a series of antimycotic compounds have been shown to demonstrate a potent activity against a wide spectrum of insect pathogenic fungi. All the preparations examined appeared to be active against isolates of *A. apis*. Chitosan in 1% salicylic acid appeared to inhibit *in vitro* the growth of the tested strains at a low concentration. At the same concentration it also revealed fungicide action (Tab.3)

Tab. 3

Effects of antimycotics on *Ascosphaera apis*, brood and worker bees
Efekt oddziaływania substancji przeciwczybowych na *Ascosphaera apis* oraz czerw i robotnice pszczoły miodnej

Antimycotics	MIC (µg/ml)	MFC (µg/ml)	LD ₅₀ (µg/insect)	
			brood	imago
<i>A. officinalis</i>	1.5 - 2.0	4.5 - 5.0	145.0	125.0
<i>P. sativa</i>	35.0 - 38.0	41.0 - 49.0	310.0	280.0
<i>L. intermedia</i>	25.0 - 40.0	400.0	105.0	80.0
Nystatin	328.0 - 340.0	350.0	128.0	93.0
Choline salt of N-glucosyl-polifungine	5.5 - 15.0	10.0 - 20.0	320.0	210.0
Chitosan + salicylic acid (Chitosal)	18.7		> 2000.0	> 2300.0

The test methods used to evaluate the toxicity and the hazard of the testing preparations to brood and worker bees showed that Chitosal-Apis® Liquid is virtually non toxic to brood and bees (tab.3). After a continuous

feeding of the bee larvae the value of the LD₅₀ exceeded 2000 and for worker bees it was more than 2300 µg per insect. These values of LD₅₀ were many times higher than those noted for furanocoumarin extracts, Nystatin and choline salt of N-glucosyl polifungine.

Preliminary observations in the field showed that Chitosal-Apis[®] Liquid used in the spring efficiently controls chalkbrood (Fig.1.). The liquid was found to have no effect in only 10% of the colonies and a recurrence appeared next spring in only 8% of the colonies in which symptoms of chalkbrood disappeared after the treatment in the previous year. Autumn treatment was much more effective. The signs of chalkbrood disappeared completely after just two applications of the preparation under study. Moreover, in only 2% of the treated colonies did chalkbrood reappear in the spring next year. In non-treated colonies chalkbrood persisted for the whole period of observation.

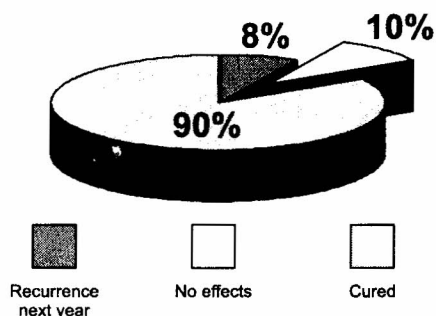


Fig.1. Chalkbrood therapy in spring with Chitosal-Apis[®] Liquid.
Efekty wiosennego leczenia grzybicy otorbielakowej preparatem Chitosal-Apis[®] plyn.

DISCUSSION

This study on the effectiveness of Chitosal-Apis[®] Liquid in controlling the natural causes of chalkbrood showed that it is possible to use this preparation in apiaries. The active ingredients of the preparation acted as fungistatic by preventing both the germination of spores and the multiplication of fungal hyphae. They also acted as a fungicidal agent because they destroyed completely the infectious factor. Moreover, Chitosal-Apis[®] Liquid appeared to be non toxic and was well tolerated by the bee colony at concentrations that effectively suppressed the invading fungus, and the preparation was easy to apply in apiaries. The *in vitro* efficacy of the examined preparation was also very high. This effectiveness against *Ascosphaera apis* was confirmed by field studies in colonies naturally infected with chalkbrood. Therefore, it can be reasonably assumed that Chitosal-Apis[®] Liquid is a potent antifungal preparation easy to apply in bee colonies. It is worthy of note that in treated

colonies the development of brood, the queen's egg laying and the behaviour of worker bees were not disturbed.

It is very interesting to note the modulation of cellular immune responses of bees by chitosan. In this way, the treatment reduces the susceptibility of the brood to *A. apis* infection caused by the depression of antiinfectious protective immunity (Gliński et al.2000)

The treatment of chalkbrood with Chitosal-Apis® Liquid was not supported by sanitary and hygienic manipulations. Hence, because of the good effects of the use of Chitosal-Apis® Liquid by itself in treated colonies, sanitary and hygienic manipulations could be avoided.

When chemicals are introduced into the bee colonies, they often contaminate the honey, the pollen loads and even the wax. As a result contamination creates a threat for consumers of honey, pollen and propolis. Thus, from an hygienic point of view, the dangers of residual chemicals and drugs in bee products to consumers are important. (Gliński 2000). Chitosan-derivatives in this respect are safe for consumers, because a) they are obtained from chitin which is a natural product commonly found in nature, b) Chitosal-Apis® Liquid is applied in apiaries in spring and in autumn, at a time when market honey has been removed from the bee hives and it cannot come to contact with this preparation.

CONCLUSIONS

1. Chitosal-Apis® Liquid is virtually non toxic for brood and worker bees both in laboratory tests and in examinations in bee colonies.
2. Chitosal-Apis® Liquid appeared to be very effective in laboratory studies and in treatment of chalkbrood in infected honey bee colonies. Bee colonies should be treated in spring or in autumn without any sanitary or hygienic manipulations (shaking, disinfection).

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**NOWA METODA ZWALCZANIA GRZYBICY
OTORBIELAKOWEJ CZERWIA PSZCZOŁY MIODNEJ
Z ZASTOSOWANIEM PREPARATU CHITOSA-APIS[®] PŁYN**

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S t r e s z c z e n i e

Mechanizm działania większości substancji o działaniu przeciwgrzybiczym polega na zaburzeniu biosyntezy ergosterolu w komórce grzyba. Pochodne chityny, które cechuje aktywność przeciwgrzybicza, najprawdopodobniej powodują zmiany w przepuszczalności błon strzępki grzyba, czego efektem jest zahamowanie wzrostu oraz śmierć komórki. Chitosal-Apis[®] Płyn, który jest mieszaniną 2-acetamido-2-deoxy-β-D-glucose and 2-amino-deoxy-β-glucopyranose w 1% roztworze kwasu salicylowego, nie tylko hamuje

wzrost grzyba, ale powoduje śmierć komórki *Ascospaera apis* -czynnika etiologicznego grzybicy otorbielakowej czerwia pszczoły miodnej.

Preparat jest nietoksyczny dla czerwia i pszczół. Bardzo dobre efekty terapeutyczne uzyskano stosując Chitosal-Apis® Płyn na wiosną w formie oprysku czerwia i pszczół oraz w jesieni w formie dokarmiania na zimę. Zarówno w trakcie leczenia jak i po jego zakończeniu nie obserwowano niepożądanych efektów. Ze względu na fakt, że Chitosal-Apis® Płyn zawiera jako substancje czynne produkty pochodzenia naturalnego (pochodne chityny, kwas salicylowy) dawka lecznicza jest mała oraz leczenie jest przeprowadzane po odwirowaniu miodu towarowego, produkty pszczele pochodzące z leczonych rodzin nie stanowią zagrożenia dla zdrowia konsumentów.

Słowa kluczowe: Chitosal-Apis® Płyn, grzybica otorbielakowa, pszczoła miodna.